

AN EASY AND RELIABLE METHOD FOR PAH EXTRACTION FROM FOOD SAMPLES

Naghiu¹ A., D. Simedru², C. Laslo¹, A. Mihaltan²

¹ University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca, 400372 Cluj-Napoca, Romania

² INCDO-INOE2000, Research Institute for Analytical Instrumentation, ICIA Cluj-Napoca Subsidiary, 400293 Cluj-Napoca, Romania

Abstract. *A simple and reliable method for the determination of 16 PAHs from meat and meat products is introduced. The method uses just 10 g of sample and has a high recovery of 70-85%. Prior to the liquid/liquid extraction with cyclohexane the meat samples are saponified using an alcoholic solution of KOH. The samples were purified using a Florosil column and analyzed with a HPLC-FLD instrument. The method was tested on sever meat products that are found on the Cluj-Napoca market with excellent results.*

Keywords: PAHs, HPLC-FLD, extraction method, meat and meat products

INTRODUCTION

Polycyclic aromatic hydrocarbons (PAHs) are a group of about 10,000 organic compounds containing two or more fused aromatic rings that are formed and released during incomplete combustion or pyrolysis of organic matter, during industrial processes and other human activities.[1,2]

Compounds that are relevant considering their effect on human health and there abundance in the environment are naphthalene, acenaphthylene, acenaphthene, fluorene, phenanthrene, anthracene, fluoranthene, pyrene, benz[a]anthracene, chrysene, benzo[b]fluoranthene, benzo[k]fluoranthene, benzo[a]pyrene, indeno[1,2,3,c,d]-pyrene, dibenz[a,h]anthracene, benzo[g,h,i] perylene [2, 3, 4, 5].

The most studied PAH is benzo[a]pyrene, which is often used as a marker for PAH in ambient air and food.[2]

Human's major routes of exposure to PAH are from inhaled polluted air and food products, especially the ones that suffer grilling, roasting and smoking processes. [2, 6, 7, 8] Vegetables may also contain high values of PAHs due to the air pollution and the deposit of these organic pollutants on the surface. [2, 6, 7]. PAHs are lipophilic compound and thus tend to form complex bound with the fatty part of meat products. [1, 9]

Considering the complexity of the sample matrix and the low concentration in which the analyte is found, in order to have a sensitive, selective and stable method of analysis it is essential to have an easy, reliable and rugged method of extraction.

MATERIAL AND METHOD

Reagents and standards. PAH Calibration Mix containing 10µg/ml of each compound (Naphthalene, Acenaphthene, Fluorene, Phenanthrene, Anthracene, Fluoranthene, Pyrene, Benz[a]anthracene, Chrysene, Benzo[b]fluoranthene, Benzo[k]fluoranthene, Benzo[a]pyrene, Dibenz[a,h]anthracene, Benzo[ghi]perylene, Indeno[1,2,3-cd]pyrene) in Acetonitrile was acquired from Supelco.

Cyclohexane for HPLC (purity $\geq 99.9\%$), Ethanol and potassium hydroxid, Acetonitrile Chromasolv gradient grade for HPLC (purity $\geq 99.9\%$) was acquired from Sigma – Aldrich. The ultra-pure water was obtained with a Milli-Q water purification system from Millipore.

Florisil (Merck) was used after heating overnight at 120°C . $0,45\mu\text{m}$ filtration cartridge for syringe where acquired from Phenomenex.

Samples

The extraction method was tested on 3 random choose meat products that were bought from the Cluj-Napoca market. The meat products had different fat content and different heat treatment during their production. The samples analyzed are: loin, bacon and baloney.

Liquid chromatography conditions

The method was developed using a Perkin Elmer 200 Series High Performance Liquid Chromatograph (HPLC) with UV and FLD detectors.

System Parameters:

- Flow Rate: $1,6\text{mL}/\text{min}$
- Mobile Phase: A (H_2O)
B (ACN)
- Column Temp: 25°C
- Injection Volume: $20\ \mu\text{L}$
- Column: ZORBAX Eclipse PAH $5\mu\text{m}$, 4.6×150 mm column from Agilent Technologies
- Wavelength: $254\ \text{nm}$ for the UV detector, different wavelenghts appropriate for each compound for the FLD detector

Recovery experiments and preparation of blank extracts

For the study of the recovery a $10\ \text{g}$ of meat sample was spiked with $1\ \text{ml}$ standard solution containing all the 15 PAH's in a concentration of $50\mu\text{g}/\text{ml}$ dissolved in acetonitrile. In the same time a blank sample from the same meat was analyzed in order to correctly calculate the recovery.

RESULTS AND DISCUSSION

The extraction method uses $10\ \text{g}$ of sample that is homogenize in a laboratory blander.

After the homogenization the sample goes through a saponification step in order to dissolve all the fat that the sample contains. This is a very important step that ensures a high recovery. For the saponification step $50\ \text{ml}$ of KOH solution $0,4\ \text{M}$ in ethanol and water (9:1) was used. The sample was then put in a ultrasound bath for 30 minutes at 60°C . Before the liquid/liquid extraction the sample was filter through a paper filter. The liquid/liquid extraction was done twice, using a separation funnel, each time using $15\ \text{ml}$ of cyclohexane. The supernatant was purified with a Florosil column and the evaporated to dryness in a gentle nitrogen stream. The sample was reconstituted using $1\ \text{ml}$ of acetonitrile. Before being injected the sample were filtered using a $0,45\mu\text{m}$ filtration cartridge.

20 μ L of the samples were then injected in the HPLC-FLD. The gradient program is shown in table 1 and in table 2 is shown the wavelengths program for each of the 15 PAH's that were analyzed.

Table 1.

Gradient program for PAHs separation by HPLC

No.	Time (min.)	Flow (ml/min)	Water A (%)	Acetonitrile B (%)
Step 1	1	1,6	55	45
Step 2	5	1,6	40	60
Step 3	15	1,6	10	90
Step 4	4	1,6	0	100
Step 5	2	1,6	0	100
Step 6	6	1,6	55	45
Step 7	17	1,6	55	45

The following formula was used in order to establish the recovery:

$$R = \frac{A_s - A_b}{A_a} * 100$$

where:

R – recovery percentage,

A_s – amount of compound found in spiked sample,

A_b – amount of compound in sample,

A_a – amount of compound added.

The recovery for all 15 PAHs analyzed are shown in table 3.

Table 2.

Wavelengths program for PAHs determination by HPLC

Compound	Wavelength (nm)		Time (min)	Gain*
	Excitation	Emission		
1. Naphthalene	224	330	0	3
2. Acenaphthene				
3. Fluorene				
4. Phenanthrene	254	402	9,9	3
5. Anthracene	237	440	10,9	4
6. Fluoranthene				
7. Pyrene	270	390	13,4	3
8. Benz[a]anthracene				
9. Chrysene				
10. Benzo[b]fluoranthene				
11. Benzo[k]fluoranthene				
12. Benzo[a]pyrene				
13. Dibenz[a,h]anthracene	270	390	17,6	4
14. Benzo[ghi]perylene				
15. Indeno[1,2,3-cd]pyrene	300	500	27,4	3

*Gain order ranges from 1-5 where 1 is the highest and 5 is the lowest.

Table 3

Recovery for the 15 PAHs by liquid/liquid extraction of food sample

Nr. Crt.	Name of compound	Recovery (%)
1	Naphthalene	81,2
2	Acenaphthene	75,4
3	Fluorene	73,2
4	Phenanthrene	69,8
5	Anthracene	77,9
6	Fluoranthene	73,8
7	Pyrene	71,3
8	Benz[a]anthracene	84,3
9	Chrysene	78,4
10	Benzo[b]fluoranthene	75,0
11	Benzo[k]fluoranthene	79,5
12	Benzo[a]pyrene	77,1
13	Dibenz[a,h]anthracene	75,8
14	Benzo[ghi]perylene	69,9
15	Indeno[1,2,3-cd]pyrene	84,5

The chromatogram obtained from the sample spiked with 50µg/ml standard solution containing all the 15 PAHs is shown in Figure 1.

All 3 samples of loin, bacon and baloney where prepared using the liquid/liquid extraction method and then analyzed using the HPLC-FLD instrument (Figure 2).

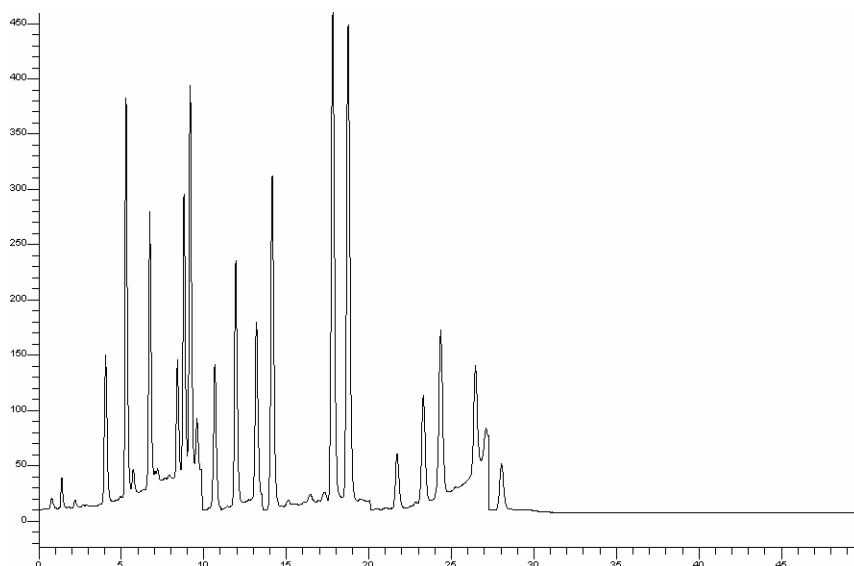


Figure 1. Chromatogram from the meat sample spiked with 50µg/ml standard solution containing all the 15 PAHs

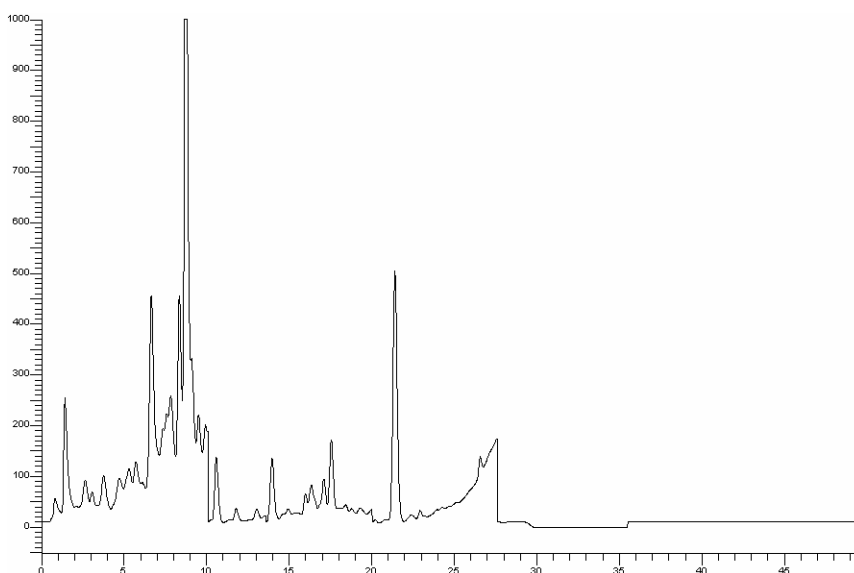


Figure 2. Chromatogram of bacon obtained using HPLC-FLD

Conclusions

The recoveries obtained with the liquid/liquid extraction are very good. It is an easy, reliable and rugged method, optimal for the extraction of meat and meat products. The amount of solvents used is low. The Florosil column retains all the fat that is left after the extraction, assuring an impurity free sample. No matrix effect was observed for the compounds of interest in the analyzed samples, they have the same retention time as the standard solution.

REFERENCES

1. ATSDR, 1995, Chemical and Physical Information, in: Toxicological Profile for Polycyclic Aromatic Hydrocarbons (PAHs), ATSDR, Atlanta, Georgia, USA, pp. 209–221 (<http://www.atsdr.cdc.gov/toxprofiles/tp69-c3.pdf>).
2. European Commission, 2002, Opinion of the Scientific Committee on Food, (http://europa.eu.int/comm/food/fs/sc/scf/out153_en.pdf).
3. Vladimír Kummer, Jarmila Mašková, Zdeněk Zralý, Martin Faldyna, 2011, Ovarian disorders in immature rats after postnatal exposure to environmental polycyclic aromatic hydrocarbons, *Journal of Applied Toxicology*, Early View (Online Version of Record published before inclusion in an issue).
4. Mi-Kyung Song, Youn-Jung Kim, Mee Song, Han-Seam Choi, Yong-Keun Park, Jae-Chun Ryu, 2011, Polycyclic aromatic hydrocarbons induce migration in human hepatocellular carcinoma cells (HepG2) through reactive oxygen species-mediated p38 MAPK signal transduction, *Cancer Science*, Vol. 102, Issue 9, pp. 1636–1644.
5. Yan-Shen Shan, Jung-Hua Fang, Ming-Derg Lai, Meng-Chi Yen, Pin-Wen Lin, Hui-Ping Hsu, Chian-Yuh Lin, Yi-Ling Chen, 2011, Establishment of an orthotopic transplantable gastric cancer animal model for studying the immunological effects of new cancer therapeutic modules, *Molecular Carcinogenesis*, Vol. 50, Issue 10, pp. 739–750.
6. Martena, M.J., M.M.P. Grutters, H.N. De Groot, E.J.M. Konings, I.M.C.M. Rietjens, 2011, Monitoring of polycyclic aromatic hydrocarbons (PAH) in food supplements containing botanicals and other ingredients on the Dutch market, *Food Additives & Contaminants: Part A*, Vol. 28, Issue. 7, pp. 925-942.
7. Martorell Isabel, Gemma Perelló, Roser Martí-Cid, Victòria Castell, Juan M. Llobet, José L. Domingo, 2010, Polycyclic aromatic hydrocarbons (PAH) in foods and estimated PAH intake by the population of Catalonia, Spain: Temporal trend, *Environment International*, Volume 36, Issue 5, pp. 424-432.
8. Stumpe-Vīksna Ilze, Vadims Bartkevičs, Agnese Kukāre, Andris Morozovs, 2008, Polycyclic aromatic hydrocarbons in meat smoked with different types of wood, *Food Chemistry*, Volume 110, Issue 3, pp. 794-797.
9. Jan H. Christensen, Giorgio Tomasi, Arthur de Lemos Scofield, Maria de Fatima Guadalupe Meniconi, 2010, A novel approach for characterization of polycyclic aromatic hydrocarbon (PAH) pollution patterns in sediments from Guanabara Bay, Rio de Janeiro, Brazil, *Environmental Pollution*, Volume 158, Issue 10, pp. 3290-3297.