

Chemical Composition and Bioactive Compounds of Some Wild Edible Mushrooms

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ABSTRACT

Recently, mushrooms consumption has risen notably thanks to the proof of their capacity to improve the organism efficiency in the combat and prevention of several diseases. The particular nutritional properties of fruiting bodies of mushrooms are one of the reasons for their consumption, besides their texture and flavour for which they are considered a delicacy. In this paper data were collected from several scientific studies with the aim to characterize the chemical composition and content of bioactive compounds (moisture, ash, total carbohydrates, total sugars, crude fat, crude protein and energy) of five edible mushrooms species: *Agaricus bisporus*, *Boletus edulis*, *Cantharellus cibarius*, *Pleurotus ostreatus*, *Lactarius piperatus*.

Keywords: *bioactive compounds, chemical composition, mushrooms.*

INTRODUCTION

The term "mushroom" describes the reproductive structure of fruiting body of a fungus (Berch *et al.*, 2007). Mushrooms belong to the kingdom of fungi, a group very distinct from plants, animals and bacteria. Mushrooms often live in symbiosis with other plants, mostly the roots of certain trees, both profiting from this relationship (Olumide, 2007). Commercial mushrooms are either produced in cultivation or harvested from the wild, including forests. The species produced in cultivation are all decomposers (or saprobes) capable of completing their life cycles on dead organic matter. Most of the forest harvested mushrooms are ectomycorrhizal and can form fruiting bodies only when growing with living host trees fungus (Berch *et al.*, 2007).

Generally, mushrooms possess four functionalities, including nutritional values, tasty properties, physiological effects, and cultural characteris-

tics (Beluhan and Ranogajec, 2011). Wild growing mushrooms are known as a delicacy in many countries, due to the high proteins and trace minerals content (Kalac 2012, Murugkar and Subbulakshmi, 2005). On the other hand wild mushrooms are reported to have a high nutritional value and be a rich source of vitamins B and D (Diez and Alvarez, 2001), which lead to the increase of their contribution to our diet (Olumide, 2007). They are used in the treatment and prevention of diseases and they exhibit varied biological properties such as antibacterial, antimutagenic, antitumoral and antiviral activities (Colak *et al.*, 2009). The most widely distributed molecules with antitumor properties in mushrooms are sesquiterpenes, triterpenoids, glucans and glycoproteins (Reis *et al.*, 2011). Traditionally, people consume mushrooms also for medicinal reasons such as reducing obesity and lowering

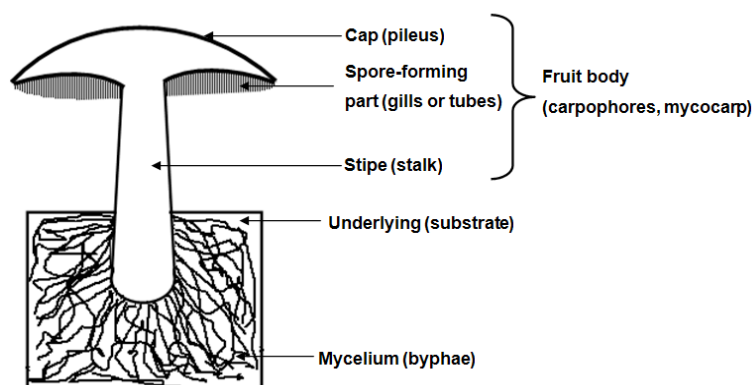


Fig. 1. General presentation of mushrooms

blood pressure in hypertensive patients (Obodai *et al.*, 2014, Armao *et al.*, 2001).

Different functional compounds have been found in mushrooms, including phenolic compounds, sterols, terpenes, ceramides, etc. (Vamanu and Nita, 2014).

Besides macronutrients with a well-balanced proportion, the studied wild mushrooms have also important micronutrients (vitamins) and non-nutrients (phenolics) with bioactive properties such as antioxidant potential (Obodai *et al.*, 2014). Among the antioxidant compounds, polyphenols and tocopherols have gained importance due to their large array of biological actions that include free radical scavenging, metal chelation enzyme modulation activities and inhibition of LDL oxidation (Keleş *et al.*, 2011, Heleno *et al.*, 2015). The high proteins and trace minerals concentration is the main reason for the significant increase of wild edible mushrooms consumption (Ogundana and Fagade, 1982, Senatore, 1990, Thimmel and Kluthe, 1998). Many researches revealed that the amino acid intake through the compositions of mushrooms are comparable to animal proteins (Fink and Hoppenhause, 1958; Gruen and Wong, 1982), which has great importance due to the fact that human nutrition has become more complex since the appearance of several diseases affecting animal meat production. However, it is evident that the step by step substitution of meat with mushroom will involve a very detailed assessment of chemical and biological properties.

Moreover, many authors have been noticed that mushrooms were used as food flavouring materials because of their unique and subtle flavor. Particularly, oct-1-en-3-ol, octan-3-ol, octan-3-on

and oct-1-en-3-on are well-known as the cause of characteristic mushroom odor (Chang and Miles, 2004; Fernandes *et al.*, 2013).

Many mushroom species and varieties are known, but the most important of these are *Agaricus spp.*, *Boletus spp.*, *Pleurotus spp.*, *Cantharellus spp.*, and *Lactarius spp.*

CHEMICAL COMPOSITION OF MUSHROOM SPECIES

Research on fungi composition assessment revealed surprising results in terms of variety of classes of compounds identified in their quantity. It is known that the chemical composition of the fungi, and thus the nutritional value depends on various factors: the variety of fungi, the soil in which the plant is growing, soil and climate maturation stage (Mattila *et al.*, 2002, Stojkovic *et al.*, 2014).

The higher fungi and the parts above ground such as the fruiting body are called mushrooms (carpophore, mycocarp). A fruiting body is formed from spacious underground mycelia (hyphae) by the process of fructification and has a lifespan generally between 10 and 14 days. The main description of the fruiting body is shown in Fig. 1. Mycelia of ectomycorrhizal species live in symbiosis with roots of a plant, mostly a tree. Terrestrial saprobic species gain nutrients mostly from organic compounds of the plant and animal debris.

Overall considerations

Several authors have reported significant specific variations in the nutritive compounds in different species of mushrooms, origin from Portugal, Croatian, Serbia (Beluhan and Ranogajec,

Tab. 1. Proximate chemical composition of wild edible mushroom species in a dry weight basis.

Species	Crude protein	Lipids	Ash	Carbohydrates	Reference
<i>Agaricus bisporus</i>	38.90	2.70	3.50	37.50	Beluhan and Ranogajec, 2011
	56.30	2.70	3.50	37.50	Barros . <i>et al.</i> , 2007
	10.00	3.12	15.00	71.86	Glamoclija . <i>et al.</i> , 2015
	34.68	2.41	7.24	50.32	Nagy ., 2016
<i>Boletus edulis</i>	36.90	2.92	5.30	1.488	Beluhan and Ranogajec, 2011
	10.65	2.23	5.26	81.86	Heleno. <i>et al.</i> , 2015
	36.24	1.92	8.38	46.23	Nagy, 2016
	29.70	3.10	5.30	51.70	Cheung Peter <i>et al.</i> , 2013
<i>Cantharellus cibarius</i>	53.70	2.89	11.50	31.90	Barros <i>et al.</i> , 2008
	21.03	2.17	9.57	57.96	Nagy, 2016
	30.91	1.90	8.80	52.50	Beluhan and Ranogajec, 2011
	21.50	5.00	8.60	64.90	Cheung <i>et al.</i> , 2003
<i>Pleurotus ostreatus</i>	30.30	1.10	13.20	-	Akyüz and Kirbağ, 2010
	17.92	1.26	11.11	62.45	Nagy, 2016
	30.40	2.20	9.80	57.60	Cheung <i>et al.</i> , 2003
<i>Lactarius piperatus</i>	29.80	2.20	5.10	62.90	Barros <i>et al.</i> , 2007
	31.81	2.69	8.30	43.00	Nagy, 2016
	13.06	2.00	7.21	77.68	Vieira <i>et al.</i> , 2014

2011, Glamoclija *et al.*, 2015, Kalac, 2009). The Romanian researchers also analyzed the chemical composition of edible mushrooms from five species: fresh material and air-dried sample, harvested in Transylvania (Nagy, 2016, Vamanu and Nita, 2013). Proximate chemical composition collected of the most important mushroom species (*Agaricus bisporus*, *Pleurotus ostreatus*, *Cantharellus cibarius*, *Boletus edulis* and *Lactarius piperatus*) are given in (Tab. 1).

The chemical composition of 5 wild edible mushrooms, including moisture, ash, total carbohydrates, crude fat and crude protein were determined according to AOAC procedures. Carbohydrates content is calculated as [100 - (moisture + crude protein + lipids + ash)] (Fernandes *et al.*, 2015).

Considerable differences in the composition are evident not only among species but also within the species. Such differences can be partially explained by varied stage of fruit bodies maturity and also by the geographical origin. The data of Table 1 should be thus interpreted as a general information.

Carbohydrates and crude proteins are the two main components. Considerable differences are apparent for three species (*Agaricus bisporus*,

Boletus edulis and *Cantharellus cibarius*) reported by different authors. Low fat content (media only 2.79 g 100g⁻¹ DM) results in a reduced amount of energy (1552 kJ median 100g⁻¹ DM). If the median is expressed in a "standard mushroom" containing 100 g DM / kg, then 100 g of fresh mushrooms has an energy content of 155.2 kJ or 37.2 kcal. Thus, we can express that mushrooms are a delicacy due to low energy value (Stojkovi *et al.*, 2014).

Among other particularities the digestibility and bioavailability of mushroom parts have not been highlighted in the description of mushroom nutritional value. It is well known that the availability of mushroom constituents is limited by the high concentration of indigestible chitin (Grangeia *et al.*, 2011). Still in the case of wild growing mushrooms there is a deficiency of credible data considering that most of the literature data is focused on fresh mushrooms. This is also true for the impact of preservation methods, under storage and during different cooking processes, on the modification of the individual constituents of mushrooms.

Proteins and amino acids in mushrooms

Regarding the structure of proteins, essential amino acids, compared to the structure of the

Tab. 2. Content of free indispensable amino acids (mg / 100g DW) in five mushroom species

Species	Val	Leu	Ile	Thr	Met	Lys	Phe	Trp	Reference
<i>Boletus edulis</i>	45	43	29	95	-	52	57	72	Ribeiro <i>et al.</i> , 2008
<i>Cantharellus cibarius</i>	ND	14	10	18	-	36	10	15	Ribeiro <i>et al.</i> , 2006
<i>Pleurotus ostreatus</i>	121	29	ND	70	12	465	9	1	Beluhan and Ranogajec, 2011
<i>Amanita rubescens</i>	16	50	25	48	-	16	29	18	Ribeiro <i>et al.</i> , 2008
<i>Agaricus bisporus</i>	119	24	41	79	71	53	28	8	Beluhan and Ranogajec, 2011
<i>Lactarius piperatus</i>	42	37	24	89	-	48	53	68	Beluhan and Ranogajec, 2011

Val-valine, Leu-leucine; Ile-isoleucine; Thr-threonine; Met-methionine; Lys-lysine; Phe-phenylalanine; Trp-tryptophan; ND-not detected

Tab. 3. Proportion of major fatty acids (% of total fatty acids) in mushrooms

Species	Palmitic acid	Stearic acid	Oleic acid	Linoleic acid	Linolenic acid	Reference
<i>Agaricus bisporus</i>	10.0	4.1	1.3	75.7	0.1	Barros <i>et al.</i> , 2008
<i>Boletus edulis</i>	10.0	2.8	39.7	44.3	0.1	Barros <i>et al.</i> , 2008
<i>Cantharellus cibarius</i>	13.1	6.5	10.8	53.6	0.1	Barros <i>et al.</i> , 2008
<i>Pleurotus ostreatus</i>	8.3	2.1	35.9	41.3	0.1	Barros <i>et al.</i> , 2008
<i>Lactarius piperatus</i>	1.10	81.46	6.58	6.66	0.04	Barros <i>et al.</i> , 2007

contents of albumin, casein, gliadin are similar. Consistent with meat protein daily consumption of 200 g dried mushrooms have the effect of replacing the meat, ensuring a balance of protein. An amount of 1 kg dried mushrooms comes with an intake of two times greater than the contents of albumin beef, three times than pork and eleven times than milk. This super food, acceptance resulting in significant value bioactive compounds, stands out in a balanced diet given the absence of fats and low calorific value (Nagy, 2016). Free indispensable amino acids content (mg/100g DW) in five mushroom species are given in (Tab. 2).

The composition of mushroom proteins seems to be of a higher nutritional value than that of most plant proteins (Belitz *et al.*, 2009). The highest contents of proteins 53.7, 38.9 and 36.9 were observed in *Cantharellus cibarius*, *Agaricus bisporus* and *Boletus edulis*, respectively, while the lowest value of only 17.92% was in the *Pleurotus ostreatus*. Beluhan and Ranogajec (2011) and Barros *et al.* (2008) reported this data about the selected mushrooms. Beluhan and Ranogajec, (2011) have found that the main constituents of the amino acid were valine and lysine represented by the species, *Agaricus bisporus* and *Pleurotus ostreatus*.

Lipids in mushrooms

The content of total lipids ranges mostly from 1 g/100 DW to 4 g/100 DW (Tab. 1 and Tab. 3). Within fatty acid composition (Tab. 3), polyunsaturated linoleic acid (C_{18:2n-6}), mono-unsaturated oleic acid (C_{18:1n-9}) and nutritionally undesirable saturated palmitic acid (C_{16:0}) prevail. However, low proportion of linoleic acid seems to be characteristic for the all species of mushrooms and in a limited extent. Barros *et al.* (2011) in several researches has reported a high concentration of stearic acid represented by *Lactarius piperatus*. The occurrence of trans fatty acids in mushrooms has not been reported and it is not expected. It is evident that the low total lipid content and a low proportion of desirable n3 fatty acids limit the nutritional value of wild growing mushroom.

Major minerals in mushrooms

Many studies can be found in the literature, since the 1970s, which quantify the element contents in wild growing mushrooms. Several papers and reviews with numerous references give important information about mineral content of mushrooms (Kalac, 2012, Akyuz and Kirbag, 2010). Based on the literature, the general compo-

Tab. 4. Content of mineral element in mushroom species (% of dry matter)

<i>Element</i>	<i>Content</i>
Sodium	0.01 – 0.04
Potassium	2.0 – 4.0
Calcium	0.01 – 0.05
Magnesium	0.08 – 0.18
Phosphor	0.5 – 1.0
Sulfur	0.1 – 0.3

Tab. 5. Total phenolics (TP) and antioxidant activity (ABTS) of mushroom extracts (Keles *et al.* 2011, Robaszkiewicz *et al.*, 2010, Yim *et al.*, 2010)

Mushroom species	TP mg GAE/100 g DW)	ABTS uM Trolox/g DW
<i>Agaricus bisporus</i>	402.00	8.00
<i>Pleurotus ostreatus</i>	545.52	9.75
<i>Cantharellus cibarius</i>	120.30	23.93
<i>Boletus edulis</i>	1277.5	12.41
<i>Lactarius piperatus</i>	334.22	70.10

sitions regarding the content of mineral elements are given in (Tab. 4).

Potassium is the prevailing element followed by phosphorus, while calcium and sodium are at the opposite end, however, exceptions do exist. For instance, calcium content was 0.07-0.52 % of DM in seven species of *Agaricus* (Gursoy *et al.*, 2009). According to researchers the potassium content is not distributed evenly within fruit bodies and descends in the following order Cap-stipe-spore-forming part-spores. It was also found that potassium content in the fruit bodies are more than 20 to 40 times higher than in the parts found in the soil (Zeng *et al.*, 2012) The same tendency can be noticed in the case of phosphorus while the bioaccumulation of sodium and calcium was not been detected. Magnesium contents in fruit bodies are even lower than those in surface, mostly organic horizons of soils, from which mycelium uptakes the nutrients (Smolskaitė *et al.*, 2015).

Overall, ash content of mushrooms is somewhat higher or comparable with most of vegetables.

Antioxidant properties and phenolic profile of the wild edible mushrooms

Numerous studies report that the antioxidant activity of fungi is closely related to their poly-

phenol content (Reis *et al.*, 2012, Smolskaite *et al.*, 2015).

Several authors evaluated the antioxidant activity of mushroom extracts sequentially isolated by cyclohexane, dichloromethane, ethanol, methanol, and water from *Agaricus bisporus*, *Pleurotus ostreatus*, *Boletus edulis*, *Cantharella cibarius* and *Lactarius piperatus* (Barros L, *et al.*, 2008, Keles A. *et al.*, 2011) and were assessed by DPPH, ABTS, ferric reducing antioxidant power (FRAP), oxygen radical absorbance capacity (ORAC) methods. To evaluate the total phenolic content (TPC) of the selected species were performed Folin-Ciocalteu (Arnao *et al.*, 2001).

Recent published data (Tab. 5) revealed that total polyphenols for methanolic extract of *Pleurotus ostreatus* contain 545.52 ± 3.92 mg GAE/100g, DW (Robaszkiewicz *et al.*, 2010, Yim *et al.*, 2010). Keles *et al.* (2011) also found that the methanolic extract of *Agaricus bisporus* contains 402 mg/100 g, but they found higher value for *Boletus edulis* and *Lactarius piperatus* (1277.5 mg/100 g, 334.22 mg/100 g, respectively).

The ability to scavenge the ABTS cation radical depends on the harvesting stage and climate soil (Heleno *et al.*, 2012, Smolskaite *et al.*, 2015).

Tab. 6. Main eight-carbon volatile compounds present in mushrooms (Combet *et al.*, 2006)

Compound	Relative concentration (%)	Threshold value (ppm)	Aroma
1-Octen-3-ol	33	0.010	Mushroom like, sweet
1-Octen-3-one	0.02	0.004	Boiled mushroom
3-Octanol	1	0.018	Cod liver oil, nutty, sweet
3-Octanone	4	0.050	Sweet, fruity, musty, lavender
Octanol	0.3	0.48	Detergent, soap, orange like

Natural flavor and aroma compounds in mushrooms

Mushrooms have long been used as food or food flavouring materials because of their unique and subtle flavour which has been studied by many authors (Zawirska-Wojtasiak, 2004). Recently many papers have focused on the assessment of volatile compounds content of mushrooms, using combined gas chromatography and mass spectrometry. Thomas (1973) reported more than 60 compounds in dried fruit bodies of *Boletus edulis* among which nine pyrazines, seven 2-formyl-pyrroles, six fatty acids (C₅ to C₁₀), five furan derivatives, four lactones and four aliphatic ketones. However it is believed that some of the compounds may have been the product of the drying process. In fresh *Boletus edulis* Dudareva (1975) found benzaldehyde, 3-heptanone, isovaleric aldehyde, acetaldehyde, methyl cyclohexanone and four unknown compounds. Ethylene together with low boiling volatiles (acetaldehyde, acetone, ethanol and ethyl acetate) and short-chain fatty acids (acetic, isobutyric, isovaleric and n-butyric acids) was also identified in *Agaricus bisporus*.

As a complementary study to the composition evaluation of *Agaricus bisporus*, we intended to perform a quantitative analysis and an assessment using a taste panel which showed the impact on the flavour of the known constituents of *Agaricus bisporus*.

As a result, the parts of the mushrooms were submitted to chemical analysis, followed by an organoleptic evaluation. Also, we used as reference some flavour compounds found in other edible mushrooms. The main odorants of the mushroom (Tab. 6.) aroma are eight carbon atom (C₈) compounds including 1-octanol, 3-octanol, 3-octanone, 1-octen-3-ol, 2-octen-1-ol, and 1-octen-3-one. 1-octen-3-ol is the key contributor to mushroom flavor and have been described

by many in *Agaricus bisporus* and other fungi (Dudareva 1975, Chambers *et al.*, 1998, Combet *et al.*, 2006).

As it is well known the volatile compound are characterized by a particular odor or aroma, such as 1-octen-3-ol for sweet and ketone 3-octanone responsible for fruity. It was also found that benzyl alcohol leads to an almond-like aroma while cyclo-octanol gives a leafy odor and can result from the cyclization of 1-octen-3-ol. In addition, studies revealed that 1-octen-3-ol concentration can determine the resulting odor sensation (Jong and Birmingham, 2005, Venkateshwarlu *et al.*, 1999). There are also studies which do not always show accurate flavor profile of volatile species emitted from fruit bodies or mycelium due to the fact that they involve analytical methods, like solvent extraction, which can lead to degradation of the sample in parallel with the production of the volatile compounds concentrate.

CONCLUSIONS

Mushrooms can be used as source of alternative food in addition to fortification or supplementation of diet for enhanced nutrition, thanks to their high content of bioactive compounds. The most studied mushrooms were *Agaricus bisporus* and *Boletus edulis* in terms of chemical composition and bioactive compounds.

Studied mushrooms proved to be rich sources of proteins, carbohydrates and ash containing also different bioactive compounds such as flavonoid compounds, phenolic compounds and antioxidant activity. Polyunsaturated fatty acids predominated over mono and unsaturated fatty acids. The studied mushroom species are poor in fat content, making them low caloric foods. Furthermore, all these samples revealed antioxidant activity, being *B. edulis* more effective in ABTS and reducing power assays.

REFERENCES

1. Murugkar AD, Subbulakshmi G (2005). Nutritional value of edible wild mushrooms collected from the Khasi hills of Meghalaya. *Food Chem* 89:599–603.
2. Akyüz M, Kirbağ S (2010). Nutritive value of wild edible and cultured mushrooms. *Turkish Journal of Biology* 34:97-102.
3. Arnao MB, Cano A, Alcolea JF, Acosta M (2001). Estimation of free radical-quenching activity of leaf pigment extracts. *Phytochem Anal* 12:138-143.
4. Barros L, Baptista P, Correia D, Casal S, Oliveira B, Ferreira I (2007). Fatty acid and sugar compositions and nutritional value of five wild edible mushrooms from Northeast Portugal. *Food Chem* 105:140-145.
5. Barros L, Baptista P, Correia D, Morais JS, Ferreira IC (2007). Effects of conservation treatment and cooking on the chemical composition and antioxidant activity of Portuguese wild edible mushrooms. *J Agric Food Chem* 55:4781–4788.
6. Barros L, Baptista P, Estevinho LM, Ferreira, IC (2007). Effect of fruiting body maturity stage on chemical composition and antimicrobial activity of *Lactarius* sp. Mushrooms. *J Agric Food Chem* 55:8766–8771.
7. Barros L, Baptista P, Ferreira IC (2007). Effect of *Lactarius piperatus* fruiting body maturity stage on antioxidant activity measured by several biochemical assays. *Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association* 45:1731-1737.
8. Barros L, Cruz T, Baptista P, Estevinho LM, Ferreira IC (2008). Wild and commercial mushrooms as source of nutrients and nutraceuticals. *Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association* 46:2742-2747.
9. Barros L, Duenas M, Ferreira IC, Baptista P, Santos-Buelga C (2009). Phenolic acids determination by HPLC-DAD-ESI/MS in sixteen different Portuguese wild mushrooms species. *Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association*, 47:1076-1079.
10. Belitz HD, Gursoy N, Sarikurkcu C, Cengiz M, Solak M.H (2009). Antioxidant activities, metal contents, total phenolics and flavonoids of seven *Morchella* species. *Food Chem Toxicol* 47:2381-2388.
11. Beluhan S, Ranogajec A (2011). Chemical composition and non-volatile components of Croatian wild edible mushrooms. *Food Chem* 124:1076-1082.
12. Berch SM, Ka KH, Park H, Winder R (2007). Development and potential of the cultivated and wild-harvested mushroom industries in the Republic of Korea and British Columbia. *Journal of Ecosystems and Management* 8 (3):53-75.
13. Chambers IVE, Smithz EC, Seitz LM, Sauer DB (1998). Sensory properties of musty compounds in food. In: *Food flavors: formation, analysis and packaging influences*. Eds E.T. Contis, C.T. Ho, C.J. Mussinan, T.H. Parliament, F. Shahidi, A.M. Spanier. Elsevier Amaster, 173-180.
14. Chang ST, Miles PG (2004). *Mushrooms: cultivation, nutritional value, medicinal effect, and environmental impact*. 2nd ed.; Boca Raton, FL, USA: CRC Press.
15. Chernov N, Osolina S, Nikitina A (2013). Chemical composition of *agaricus bisporus* and *pleurotus ostreatus* fruiting bodies and their morphological parts. *Journal of Faculty of Food Engineering, Romania Volume XII, Issue 4:291 – 299*.
16. Cheung LM, Cheung PCK, Ooi VEC, (2003). Antioxidant activity and total phenolics of edible mushroom extracts. *Food Chem* 81:249–255.
17. Colak A, Faiz Ö, Sesli E (2009). Nutritional composition of some wild edible mushrooms. *Turkish Journal of Biochemistry* 34:25–31.
18. Diez VA, Alvarez A (2001). Compositional and nutritional studies on two wild edible mushrooms from northwest Spain. *Food Chem* 75:417-422.
19. Dudareva NT (1975). Aromatic substances of fresh and sublimation-dried mushrooms. *Priki Biokhim Mikrobiol* 11:147-148.
20. Fernandes A, Barreira JCM, António AL, Santos PMP, Martins A, Oliveira MBPP, Ferreira ICFR (2013). Study of chemical changes and antioxidant activity variation induced by gamma-irradiation on wild mushrooms: comparative study through principal component analysis. *Food Res Int* 54:18–25.
21. Fink H, Hoppenhaus KW (1958). Peculiar observations in the estimation of biological quality of the proteins of edible boletus (*Boletus edulis*) and mushroom (*Psalliota biospora*) with reference to dietetics and therapeutics. *Nutrition Abstracts Reviews* 28:4886.
22. Glamoclija J, Stojkovic D, Nikolic M, Ciric A, Reis FS, Barros L, Ferreira IC, Sokovic M, (2015). A comparative study on edible *Agaricus* mushrooms as functional foods. *Food & function* 6:1900-1910.
23. Grangeia C, Sandrina AH, Barros L, Martins A, Ferreira ICFR (2011). Effects of trophism on nutritional and nutraceutical potential of wild edible mushrooms. *Food Res Int* 44:1029–1035.
24. Gruen EH, Wong MW (1982). Distribution of cellular amino acids, protein and total inorganic nitrogen during fruit body development in *Flammulina velutipes*. *Canadian Journal of Botany* 60:1330–1341.
25. Heleno SA, Barros L, Martins A, Morales P, Fernández-Ruiz V, Glamoclija J, Sokovic M, Ferreira ICFR, (2015). Nutritional value, bioactive compounds, antimicrobial activity and bioaccessibility studies with wild edible mushrooms. *LWT - Food Sci Techn* 63:799-806.
26. Jong SC, Birmingham JM (2005). Mushrooms as a source of natural flavor and aroma compounds. *American type culture collection*, 37:345-366.
27. Kalač P (2009). Chemical composition and nutritional value of European species of wild growing mushrooms. A review. *Food Chem* 113:9-16.
28. Kalac P (2012). Chemical composition and nutritional value of European species of wild growing mushrooms. Nova Science Publisher, In S. Andres & N. Baumann (Eds.), *Mushrooms: Types, properties and nutrition* pp. 130–151.

29. Keles I, Koca, Gencelep H (2011). Antioxidant properties of wild edible mushrooms. *Journal of Food Processing Technology*, 2:130.
30. Mattila P, Salo-Väänänen P, Könkö K, Aro H, Jalava T (2002). Basic composition and amino acid contents of mushrooms cultivated in Finland. *J Agric Food Chem* 50:6419–6422.
31. Nagy M (2016). Research regarding the upper valorisation of some vegetable sources rich in bioactive compounds in order to obtain an innovative meat product. Phd Thesis.
32. Nobukuni T, Narida K, Yamauchi K (1970). Food seasonings with mushroom flavor. Japanese Patent 70 23,580; Chem Abstr 74 ref 63330f.
33. Obodai M, Ferreira IC, Fernandes A, Barros L, Mensah DL, Dzomeku M, Urben AF, Prempeh J, Takli RK (2014). Evaluation of the chemical and antioxidant properties of wild and cultivated mushrooms of Ghana. *Molecules* 19:19532-19548.
34. Ogundana SK, Fagade OE (1982). Nutritive value of some Nigerian edible mushrooms. *Food Chem* 8:263–268.
35. Olumide OJ (2007). Economic analysis of mushroom marketing as a coping strategy for poverty reduction in Ondo state Nigeria. In *Proceeding of African Crop Science Conference, El-Minia Egypt* 8:1255-1260.
36. Reis FS, Martins A, Barros L, Ferreira IC, (2012). Antioxidant properties and phenolic profile of the most widely appreciated cultivated mushrooms: a comparative study between in vivo and in vitro samples. *Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association*, 50:1201-1207.
37. Reis FS, Pereira E, Barros L, Sousa MJ, Martins A, Ferreira IC (2011). Biomolecule profiles in inedible wild mushrooms with antioxidant value. *Molecules*, 16:4328-4338.
38. Ribeiro B, Rangel J, Valentão P, Baptista P, Seabra RM, Andrade PB (2006). Contents of carboxylic acids and two phenolics and antioxidant activity of dried Portuguese wild edible mushrooms. *Journal of Agricultural and Food Chem* 54:8530–8537.
39. Ribeiro B, Lopes R, Seabra RM, Gonçalves RF, Baptista P, Quelhas I (2008). Comparative study of phytochemicals and antioxidant potential of wild edible mushroom cap and stipes. *Food Chem* 110:47–56.
40. Robaszkiewicz A, Bartosz G, Lawryniewicz M, Soszynski M (2010). The role of polyphenols, β -carotene, and lycopene in the antioxidative action of the extracts of dried, edible mushrooms. *Journal of Nutrition and Metabolism* 42:1–9.
41. Senatore F (1990). Fatty acids and free amino acid content of some mushrooms. *J Sci Food Agric* 51:91–96.
42. Smolskaitė L, Venskutonis PR, Talou T (2015). Comprehensive evaluation of antioxidant and antimicrobial properties of different mushroom species. *LWT - Food Science and Technology* 60:462-471.
43. Stojkovi D, Reis F, Glamoclija J, Ciric A, Barros L, Van Griensven LJLD, Ferreira ICFR, Sokovic M (2014). Cultivated strains of *Agaricus bisporus* and *A. brasiliensis*: chemical characterization and evaluation of antioxidant and antimicrobial properties for the final healthy product – natural preservatives in yoghurt. *Food Funct* 5:1602–1612.
44. Svoboda L, Chrastný V (2008). Levels of eight trace elements in edible mushrooms from a rural area. *Food Additives and Contaminants* 25:51–58.
45. Thimmel R, Kluthe R (1998). The nutritional database for edible mushrooms. *Ernahrung* 22:63–65.
46. Thomas AF (1973). An analysis of the flavor of the dried mushroom, *Boletus edulis*. *J Agr Food Chem* 21:955-958.
47. Vamanu E, Nita S (2013). Antioxidant capacity and the correlation with major phenolic compounds, anthocyanin, and tocopherol content in various extracts from the wild edible *Boletus edulis* mushroom. *BioMed research international* 313-905.
48. Venkateshwarlu G, Chandravadana MV, Tewari R P (1999). Volatile flavour components of some edible mushrooms (*Basidio mycetes*). *Flavour and Fragrance Journal* 14:191–194.
49. Vieira V, Barros L, Martins AI, Ferreira CFR (2014). Expanding current knowledge on the chemical composition and antioxidant activity of the genus *Lactarius*. *Molecules* 19(12): 20650-20663.
50. Yim H S, Chye FY, Tan CT, Ng YC, Ho CW (2010). Antioxidant activities and total phenolic content of aqueous extract of *Pleurotus ostreatus* (cultivated oyster mushroom). *Malaysian Journal of Nutrition* 16:281-291.
51. Zawirska-Wojtasiak R, Siwulski M, Mildner-Szkudlarz S, Wąsowicz E (2009). Studies on the aroma of different species and strains of *pleurotus* measured by GC/MS, sensory analysis and electronic nose. *Acta Sci Pol, Technol Aliment* 8(1):47-61.